

Short Communication

A note on pathogenic variations among isolates of *Colletotrichum graminicola* causing anthracnose disease of sorghum

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Key words: Anthracnose, *C. graminicola*, pathotype, sorghum, virulence diversity

Anthracnose caused by *Colletotrichum graminicola* is one of the most devastating diseases of sorghum. The disease appears in severe form on forage sorghum under northern Indian conditions where yield losses up to 50% or more can occur (Thakur *et al.*, 2007). Use of resistant cultivars as a strategy to control the disease has often met with limited success because of the breakdown of resistance. The pathogen is highly variable and known to have physiological races (Mathur *et al.*, 2000). Pande *et al.* (1991) studied 9 isolates of *C. graminicola* from India and observed large genetic variation present in them. To develop a sustainable anthracnose resistance breeding programme, information on the present state of variation in the pathogen population is essential. There is no report of variability study in sorghum anthracnose for almost last two decades. The objective of the present investigation was to study the virulence diversity among different isolates of *C. graminicola* causing anthracnose of sorghum.

Ten cultures which included five isolates from Udham Singh nagar (UKL-Uttarakhand Mandawali isolate, UKK-Uttarakhand Kichha isolate, UKN- Uttarakhand Gadarpur isolate, UKA- Uttarakhand Pantnagar isolate, UKF-Uttarakhand Sitarganj isolate), two isolates from Haridwar (UKD-Uttarakhand Patanjali isolate, UKS-Uttarakhand Bahadarabad isolate), two isolates from DehraDun (UKB-Uttarakhand Doiwala isolate, UKC-Uttarakhand Dhakrani isolate) and one isolate from Nainital (UKH-Uttarakhand Haldwani isolate) were obtained from a repository of *C. graminicola* isolates available at sorghum pathology laboratory of the Department of Plant Pathology, GBPUA&T, Pantnagar. The cultures that showed morphological and cultural variations in the laboratory and pathogenic variations under glass house conditions were further evaluated under field conditions during two consecutive kharif seasons of 2013 and 2014 using a set of ten selected cultivars/lines. Sorghum lines grown in a plot of 2 rows of 4 m length spaced at 45x15 cm in RBD with three replications were spray inoculated with spore suspension of each isolate

@1x10⁶ conidia ml at 15-and 30 days after sowing (DAS). Observations on disease severity were taken 15 days after inoculation and at 50% flowering using a continuous 1-9 scale where: 1=<1% leaf area covered; 2=1-5% leaf area covered; 3=6-10% leaf area covered; 4=11-20% leaf area covered; 5=21-30% leaf area covered; 6=31-40% leaf area covered; 7=41-50% leaf area covered; 8=51-75% leaf area covered; 9=76-100% leaf area covered (Thakur *et al.*, 2007). Analysis of the data was done using GBPUAT Analytical Programme STPR-723 for pooled analysis.

The isolates produced typical disease symptoms on plants. On the basis of type of symptoms produced on foliage and the virulence, the isolates could be differentiated into seven categories (Table 1).

Isolates UKK, UKC and UKH were very similar in pathogenicity. At 15 DAS these isolates did not infect IRAT 204, IS 2312, CSV 21F and PC 5. These isolates produced black dot like acervuli on the foliage of Kekri local. These isolates were therefore, considered one pathotype. Isolates UKB and UKN were found to be similar in pathogenicity. These isolates were similar to isolates UKK, UKC and UKH in not infecting IS 2312 but unlike these isolates they failed to infect IS 18442 and IS 3089 at 15 DAS. Isolate UKL could not infect K local, IS 18442, IRAT 204, SSG 59-3 and PC 5 at 15 DAS but resulted in severe mid rib infection in K local and CSV 21F in addition to leaf spots at 30 DAS, thus considered as a different pathotype. Isolate UKD did not infect IS 8354 and CSV 21 F at 15 DAS. Isolate UKA like Isolate UKL could not infect K local initially but it was distinct from Isolate UKL in infecting other cultivars at 15 DAS. Isolate UKF also varied in virulence on different cultivars on inoculation at 15 DAS. Isolate UKS infected only K local, IS 3089, R local and SSG 59-3 at 15DAS and was considered a separate pathotype. However, every isolate infected all the cultivars used in the investigation to a varying extent when inoculated at 30 DAS.

In previous studies on pathogenic variability, anthracnose

Table 1: PDI and putative pathotyping of *Colletotrichum graminicola* isolates on sorghum lines

Isolates	PDI (Pooled mean of 2013 and 2014 crop season)																		Acervulus production/ midrib infection	Putative Patho- typing		
	Sorghum lines																					
	K Local	IS8354	IS18442	IRAT204	IS2312	IS3089	CSV21F	R Local	SSG59-3	PC5												
I*	II	I	II	I	II	I	II	I	II	I	II	I	II	I	II	I	II					
UKL	0.00	80.33	30.34	74.83	0.00	63.33	0.00	52.17	29.33	70.17	27.34	73.33	22.84	67.33	33.00	80.83	0.00	64.00	0.00	40.17	Severe midrib infection in K Local and CSV21 F	P1
UKK	29.00	79.33	28.33	67.33	25.50	65.50	0.00	66.00	0.00	60.17	27.50	74.17	0.00	65.17	32.00	73.00	26.67	69.67	0.00	40.00	Numerous acervuli on K Local	P2
UKD	33.17	73.50	0.00	69.67	30.84	71.17	27.17	66.17	28.34	63.50	33.50	78.33	0.00	63.17	33.83	73.67	32.50	71.17	22.84	41.17	Nil	P3
UKN	29.17	80.50	30.50	77.83	0.00	72.50	27.00	66.33	0.00	63.83	0.00	65.83	28.84	63.50	30.50	73.00	25.84	66.17	24.17	42.17	Nil/Slight	P4
UKS	33.67	77.83	0.00	73.00	0.00	70.67	0.00	61.67	0.00	65.50	32.17	78.00	0.00	60.17	34.84	73.50	33.83	62.00	0.00	39.83	Nil	P6
UKA	0.00	75.17	32.00	82.67	31.84	73.17	29.50	67.50	0.00	66.33	0.00	73.33	0.00	59.33	34.40	75.00	29.50	66.00	0.00	40.17	Nil	P5
UKF	33.50	75.00	0.00	78.87	0.00	67.17	30.33	64.67	32.50	65.00	0.00	81.00	26.34	57.50	34.84	80.83	0.00	59.67	24.50	37.83	Nil	P7
UKB	31.34	79.17	30.83	78.50	0.00	67.33	28.67	66.00	0.00	63.33	0.00	75.33	28.67	62.50	29.34	69.33	26.00	67.67	21.33	37.33	Nil/Slight	P4
UKC	27.84	77.83	30.00	79.50	30.50	67.00	0.00	61.50	0.00	66.50	35.17	73.83	0.00	65.50	36.50	71.17	27.83	62.33	0.00	37.17	Numerous acervuli on K Local	P2
UKH	34.17	74.33	27.00	78.17	28.83	67.33	0.00	64.33	0.00	63.50	30.67	73.67	0.00	67.00	33.00	70.17	23.84	59.83	0.00	36.83	Numerous acervuli on K Local	P2
CDat5%	1.75	1.65	1.75	1.74	1.35	1.60	1.49	2.89	1.03	1.47	1.38	1.44	1.17	1.29	1.92	2.27	1.97	1.59	1.71	1.36	-	-
CV	5.95	1.82	7.49	1.96	7.82	2.00	8.89	2.16	9.77	1.93	6.34	1.64	9.40	1.74	4.94	1.97	7.45	2.09	15.77	2.95	-	-

*I = PDI when inoculated 15 DAS, II = PDI when inoculated 30 DAS

virulence was studied only at 6 leaf stage (Thakur *et al.*, 1999). Mathur and Totla (2001) used three parameters for differentiating seven isolates of *C. graminicola* on six sorghum cultivars under glass house conditions and found additional parameters useful for finer differentiation. There are some reports suggesting that anthracnose symptoms do not develop on plants below 6-leaf stage i.e. 25 DAS (Nakamura, 1985; Ferreira and Casela, 1986; Thakur, 1995) probably due to high HCN contents in very young seedlings which may get diluted with plant growth. Similarly, in our studies plants at 30 DAS developed symptoms more readily but as the isolates could infect the seedlings of some cultivars at 15 DAS, it is likely that these isolates have the ability to overcome HCN in those lines. It has been demonstrated that *Gloeocercospora sorghi* causing zonate leaf spot of sorghum could overcome HCN by hydrolysing it with an enzyme Formamide hydrolyase (Myers and Fry, 1978). However, whether such a mechanism is used by these isolates of *C. graminicola* needs further investigation.

The study indicated prevalence of pathogenic variability among the isolates of *C. graminicola* which could prove valuable for designing the management strategies for the disease and to screen resistant germplasm for breeding programme.

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A note on host diversity of *Criconema* spp.

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Key words: *Criconema*, host diversity, host range, Nematode

Nematode species of the genus *Criconema* (Tylenchida: Criconemidae) are widely distributed and parasitize many plant species from very primitive orders to advanced ones. They are migratory ectoparasites and feed on root tips or along more mature roots. Reports like Rathore and Ali (2014) and Rathore (2017) reveal that most nematode species prefer feeding on plants of certain taxonomic group (s). In the present study an attempt has been made to precisely trace the host plant affinity of twenty-five *Criconema* species feeding on diverse plant species. Host species of various *Criconema* species reported by Nemaplex (2018) and others in literature were aligned with families and orders following the modern system of classification, i.e., APG IV system (2016). According to this system, angiosperms are classified in different clades and clades into different orders and subsequently into different families. Affinity of each *Criconema* species with its host plants was numerically determined by calculating General Association Index (GAI), and for a group of species in a particular clade(s) by having Specific Association Index (SAI) following the system of Rathore and Tiwari (2016). The status of nematode species was further supported by the classification of Berneys and Chapman (1994).

Association and affinity of 25 *Criconema* species presented in Table 1 revealed that 35.35 % plants were preferred in Rosids followed by monocots (25.252 %) and Asterid (20.202 %). Though, Rosids and Asterids are different clades but both possess dicotyledonous plants. The combination of two clades proves that dicotyledons

showed preference over monocots. Superrosids and Superasterids were represented by a few host plants only. However, Magnoliids and Gymnosperms substantially contributed in the host range of this nematode species. Though Rosids revealed greater preference over Asterids, the percent host families and orders were similar in number as reflected by similar SAI values. The SAI value was slightly higher for monocots that indicate stronger affinity. The same was higher for gymnosperms (0.467) in comparison to Magnoliids (0.413) (Table 1).

Perusal of taxonomic position of host species in Table 2 revealed that 68 % of *Criconema* spp. were monophagous and strictly fed on one host species. Of these, 20 % from Magnoliids were monophagous (*C. acriculum*, *C. grassiator*, *C. karacsi*, *C. magnolia*, *C. petasum*); 20 % from Rosids (*C. demani*, *C. featherensis*, *C. mangiferae*, *C. parmistum*, *C. ravidum*); 12 % from Asterids (*C. annulifer*, *C. acanum*, *C. celetum*); 12 % from monocots (*C. paucianmulatum*, *C. quasiclemani*, *C. warrenense*) and 4 % from gymnosperms (*C. neoaxestis*). Twenty-eight percent *Criconema* spp. were polyphagous and one oligophagous. GAI was 1.0 for all monophagous and oligophagous species, whereas the same was less than 1 for polyphagous species. Rosids contributed in the host range of all the polyphagous *Criconema* spp., while the association of host plants from other clades was more or less 50 %.

Maximum numbers of *Criconema* spp. were harboured by host families like Lauraceae (5), Magnoliaceae (5) in Magnoliids; Poaceae (5) in monocots; Fagaceae (4) and Rosaceae (4) in Rosids and Pinaceae (3) in gymnosperms.

Table 1: Association of *Criconema* species to different host parameters

Taxonomic clades	Host species	Host genera	Host families	Host orders	SAI
Magnoliids	10 (10.101)	10 (10.753)	10 (14.705)	9 (15.789)	0.413
Monocots	25 (25.252)	25 (26.882)	14 (20.588)	11 (19.298)	0.54
Superrosids	1 (1.010)	1 (1.075)	1 (1.470)	1 (1.754)	1
Rosids	35 (35.353)	30 (32.258)	22 (32.353)	19 (33.334)	0.52
Superasterids	3 (3.050)	3 (3.226)	3 (4.412)	1 (1.754)	0.714
Asterids	20 (20.202)	19 (20.430)	13 (19.118)	11 (19.298)	0.512
Gymnosperms	5 (5.000)	5 (5.376)	5 (7.353)	5 (8.772)	0.467

Figures in parentheses are per cent values; SAI=Specific Association Index

Table 2: Taxonomic position of host plants of *Criconema* spp.

S. No.	<i>Criconemaspp.</i>	Host species	No. of hostspecies	GAI	Status
1	<i>C. acriculum</i>	Magnoliids: Lauraceae (1) <i>Umbellularia californica</i>	1	1	Monophagous
2	<i>C. annulifer</i>	Asterids: Aquifoliaceae (1) <i>Ilex aquifolium</i>	1	1	Monophagous
3	<i>C. acanum</i>	Asterids: Asteraceae (1) <i>Solidago</i> sp.	1	1	Monophagous
4	<i>C. arkaense</i>	Monocots-Poaceae (2) <i>Arrhenatherum</i> sp., <i>Paspalum</i> sp.;Rosids: Cannabaceae (1) <i>Celtis accidentalis</i> ,Sapindaceae (1) <i>Acer saccharum</i>	4	0.6	Polyphagous
5	<i>C. celetum</i>	Asterids: Gesneriaceae (1) <i>Saintpaulia</i> sp.	1	1	Monophagous
6	<i>C. crotaloides</i>	Magnoliids: Lauraceae (1) <i>Umbelluria californica</i> ; Rosids:Rosaceae (1) <i>Rubusparviflorus</i> ; Asterids: Ericaceae (1) <i>Arctostaphylos manzanita</i> ; Gymnosperms:Pinaceae (1) <i>Pseudotsuga menziesii</i>	4	0.5	Polyphagous
7	<i>C. demani</i>	Rosids: Betulaceae (1) <i>Betula papyrifera</i>	1	1	Monophagous
8	<i>C. featherensis</i>	Rosids: Vitaceae (1) <i>Vitis californica</i>	1	1	Monophagous
9	<i>C. giardi</i>	Magnoliids: Lauraceae (1) <i>Persea americana</i> , Magnoliaceae (1) <i>Magnolia grandiflora</i> ;Rosids: Moraceae (1) <i>Ficus carica</i> , Rosaceae (1) <i>Fragaria x ananassa</i>	4	0.6	Polyphagous
10	<i>C. grassator</i>	Magnoliids: Magnoliaceae (1) <i>Liriodendron tulipifera</i>	1	1	Monophagous
11	<i>C. kavacsi</i>	Magnoliids: Lauraceae (1) <i>Umbellularia californica</i>	1	1	Monophagous
12	<i>C. magnoliae</i>	Magnoliids: Magnoliaceae (1) <i>Magnolia grandiflora</i>	1	1	Monophagous
13	<i>C. mangiferum</i>	Rosids: Anacardiaceae (1) <i>Mangifera indica</i>	1	1	Monophagous
14	<i>C. mutabile</i>	Magnoliids:Lauraceae (1) <i>Persea americana</i> , Monocots: Araceae (1) <i>Philodendron</i> sp., Arecaceae (1) <i>Palmaceae</i> sp., Asparagaceae (1) <i>Yucca</i> sp., Bromeliaceae(2) <i>Billbergia</i> sp., <i>Bromeliaceae</i> sp.. Dioscoreaceae (1) <i>Dioscorea</i> sp., Musaceae (1) <i>Musa</i> sp., Poaceae (10) <i>Arrhenatherum</i> sp., <i>Avena sativa</i> , <i>Axonopus</i> sp., <i>Bambusa</i> sp., <i>Cynodon dactylon</i> , <i>Echinochloa</i> sp., <i>Hordeum vulgare</i> , <i>Sorghum bicolor</i> , <i>Zea mays</i> , <i>Zoysia</i> p.,Typhaceae (1) <i>Typha</i> sp., Zingiberaceae (1) <i>Zingiber</i> sp.,Superasterids: Amaranthaceae (1) <i>Beta vulgaris</i> ,Cactaceae (1) <i>Cactaceae</i> sp., Nyctaginaceae (1) <i>bougainvillea</i> sp.; Rosids: Fabaceae (3) <i>Medicago sativa</i> , <i>Trifolium repens</i> , <i>Vigna unguiculata</i> , Juglandaceae (2) <i>Juglanshendsii</i> , <i>Juglans</i> sp., Malvaceae (1) <i>Gossypium hirsutum</i> , Moraceae (1) <i>Morus</i> sp., Rosaceae (8) <i>Fragariachiloensis</i> , <i>Malus sylvestris</i> , <i>Prunus domestica</i> , <i>Prunus dulcis</i> , <i>Prunus persica</i> , <i>Pyracantha</i> sp., <i>Pyrus communis</i> , <i>Rosa</i> sp., Rutaceae (2) <i>Citrus sinensis</i> , <i>Citrus</i> sp., Sapindaceae (1) <i>Acer</i> sp.; Vitaceae (1) <i>Vitis vinifera</i> Superrosids: Altingiaceae (1) <i>Liquidamber</i> sp.;Asterids: Acanthaceae (1) <i>Acanthus</i> sp., Aquifoliaceae(1) <i>Ilex</i> sp., Araliaceae (1) <i>Aralis</i> sp., Asteraceae (5) <i>Arctiumlappa</i> , <i>Baccharis</i> sp., <i>Dahlia</i> sp. <i>Tagetes erecta</i> , <i>Tagetes</i> sp., Convolvulaceae (2) <i>Dicondra</i> sp., <i>Ipomoea batatas</i> , Ericaceae (1) <i>Rhododendron</i> sp., Oleaceae (2) <i>Ligustrum</i> sp., <i>Syringea</i> sp., Solanaceae (2) <i>Nicotiana</i> sp., <i>Solanum lycopersicum</i> , Theaceae (1) <i>Camellia</i> sp.Gymnosperms: Pinaceae (1) <i>Pinussp</i>	60	0.564	Polyphagous
15	<i>C. neoaxestis</i>	Gymnosperms: Pinaceae (1) <i>Cedus lebanii</i>			
16	<i>C. paucianmulatum</i>	Monocot: Poaceae (1) <i>Zea mays</i>	1	1	Monophagous
17	<i>C. permistum</i>	Rosids: Vitaceae (1) <i>Vitis vinifera</i>	1	1	Monophagous
18	<i>C. petasum</i>	Magnoliids: Magnoliaceae (1) <i>Liriodendron tulipifera</i>	1	1	Monophagous
19	<i>C. quasiclemane</i>	Monocot: Cyperaceae (1) <i>Scirpus americanus</i>	1	1	Monophagous
20	<i>C. ravidum</i>	Rosids: Fagaceae (1) <i>Quercus</i> sp.	1	1	Monophagous
21	<i>C. sphagni</i>	Magnoliids: Magnoliaceae (1) <i>Liriodendron</i>	1	1	Monophagous

		<i>tulipifera</i> ; Monocots: Poaceae (1) <i>Arrhenatherum</i> sp.; Rosids: Fagaceae (1) <i>Quercus</i> sp.	3	0.556	Polyphagous
22	<i>C. tribule</i>	Rosids: Fagaceae (2) <i>Fagus</i> sp., <i>Quercus</i> sp.	2	1	Oligophagus
23	<i>C. vishwanathum</i>	Rosids: Rosaceae (2) <i>Prunus domestica</i> , <i>Prunus persica</i> Gymnosperms: Cupressaceae (1) <i>Juniperus oxycedrus</i>	3	0.833	Polyphagous
24	<i>C. warrenense</i>	Monocots: Poaceae (1) <i>Paspalum</i> sp.	1	1	Monophagous
25	<i>C. zantene</i>	Rosids: Fagaceae (1) <i>Quercus</i> sp.; Gymnosperms: Podocarpaceae (1) <i>Podocarpus</i> sp.	2	0.667	Polyphagous

C. mutabile parasitized maximum number of host species (Table 3).

Though *Criconeuma* spp. parasitize many varieties of host species, nevertheless they tend to prefer woody plants. To examine this issue further, all the host families except gymnosperms were aligned according to the classification of Hutchinson (1973). He classified angiosperms into monocotyledons and dicotyledons. Hutchinson divided monocotyledons into calyciferae (calyx bearers-with distinct (usually green) calyx and corolla), corolliferae (calyx and corolla are more or less similar), and glumiflorae (perianth is much more reduced or represented by lodicules), whereas dicotyledons were partitioned into Lignosae (fundamentally woody plants) and Herbaceae (fundamentally herbaceous group of plants). *Criconeuma* spp. parasitized plants from 28 families (Magnoliids, Superrosids, Rosids, Superasterids, Asterids) and according to Hutchinson's classification 21 aligned with Lignosae and 7 with Herbaceae indicating greater preference towards woody plants (75 %). It will be worthwhile to mention that among monocotyledons 50 % families had plants from Corolliferae. Family Poaceae was most dominating family in monocotyledons. Since, *Criconeuma* spp. showed greater preference towards woody plants, it is suggested that cultivated crops prone to these nematode species should be grown away from forest areas.

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Acknowledgements of financial support, advice, and other kinds of assistance should be made at the end of the paper under the heading "Acknowledgements".

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3.10.1 Journal articles

Van Ranst, E., Utami, S. R., Vanderdeelen, J., Shamshuddin, J. 2004. Surface reactivity of Andisols on volcanic ash along the Sunda arc crossing Java Island, Indonesia. *Geoderma*, 123, 193-203.

3.10.2 Unpublished work

Papers that are unpublished but have been accepted must be cited with the journal's name followed by (in press). In all other cases, reference must be made to (unpublished work) or (personal communication).

3.10.3 Books and monographs

Tertian, R., Claisse, F. 1982. Principles of Quantitative X-Ray Fluorescence Analysis. Heyden, London, 385 p.

3.10.4 Chapters from multi-author books

Wold, S., Sjöström, M. 1977. Chemometrics, Theory and Application. In: B. R. Kowalski (ed). ACS Symposium Series N° 52. American Chemical Society, Washington, DC, pp: 243-282.

3.10.5 Theses

Hassink, J. 1995. Organic matter dynamics and N mineralization in grassland soils. Doctoral thesis, Wageningen University, The Netherlands, 250 p.

3.10.6 Patents

Millcr, B.O. 1952. U.S. Patent 2542356, Dow Chemical Company; Chemical Abstracts 51 (1961) 2870.

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