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CONTENTS

Unrevealing the role of epistasis through Triple Test Cross in Indian mustard NARENDER SINGH, USHA PANT, NEHA DAHIYA, SHARAD PANDEY, A. K. PANDEY and SAMEER CHATURVEDI	330
Testing of InfoCrop model to optimize farm resources for mustard crop under <i>tarai</i> region of Uttarakhand	335
MANISHA TAMTA, RAVI KIRAN, ANIL SHUKLA, A. S. NAIN and RAJEEV RANJAN	
<i>In vitro</i> evaluation of endophytes and consortium for their plant growth promoting activities on rice seeds DAS, J., DEVI, R.K.T. and BARUAH, J.J.	342
Effect of subsurface placement of vermicompost manure on growth and yield of wheat (<i>Triticum aestivum</i> L. Var. UP 2526) ABHISHEK KUMAR and JAYANT SINGH	348
Assessment of different nutrient management approaches for grain yield, gluten content and net income of common bread wheat (<i>Triticum aestivum</i> l.) in Western Himalayan region of Uttarakhand BHAWANA RANA and HIMANSHU VERMA	359
Suitability assessment of land resources forc assava(<i>Manihot esculentus</i> L.) and yam (<i>Dioscorea spp L.</i>) cultivation in Khana LGA, Rivers State, Southern Nigeria PETER, K.D., UMWENI, A.S. and BAKARE, A.O.	367
Biophysical and biochemical characters conferring resistance against pod borers in pigeonpea PARUL DOBHAL, R. P. MAURYA, PARUL SUYAL and S.K. VERMA	375
Population dynamics of major insect pest fauna and their natural enemies in Soybean SUDHA MATHPAL, NEETA GAUR, RASHMI JOSHI and KAMAL KISHOR	385
Fumigant toxicity of some essential oils and their combinations against <i>Rhyzopertha dominica</i> (Fabricius) and <i>Sitophilus oryzae</i> (Linnaeus) NIDHI TEWARI and S. N. TIWARI	389
Long term efficacy of some essential oils against <i>Rhyzopertha dominica</i> (Fabricius) and <i>Sitophilus oryzae</i> (Linnaeus) NIDHI TEWARI and S. N. TIWARI	400
Management strategies under chemicals, liquid organic amendments and plant extracts against black scurf of potato caused by <i>Rhizoctonia solani</i> Kühn in <i>tarai</i> regions of Uttarakhand SURAJ ADHIKARI, SHAILBALA SHARMA, R. P. SINGH, SUNITA T. PANDEY and VIVEK SINGH	408
Effective management strategies against ginger rhizome rot caused by <i>Fusarium solani</i> by the application of chemicals, bioagents and Herbal <i>Kunapajala</i> in mid hills of Uttarakhand SONAM BHATT, LAXMI RAWAT and T. S. BISHT	417

Distribution and morphological characterisation of isolates of <i>Fusarium moniliforme</i> fsp. <i>subglutinans</i> causing Pokkah Boeng disease of sugarcane in different sugarcane growing areas of Udham Singh Nagar district of Uttarakhand HINA KAUSAR, BHAGYASHREE BHATT and GEETA SHARMA	429
Biointensive management of <i>Meloidogyne enterolobii</i> in tomato under glasshouse conditions SHUBHAM KUMAR, ROOPALI SHARMA, SATYA KUMAR and BHUPESH CHANDRA KABDWAL	435
Effect of pre-harvest application of eco-friendly chemicals and fruit bagging on yield and fruit quality of mango KIRAN KOTHIYAL, A. K. SINGH, K. P. SINGH and PRATIBHA	447
A valid and reliable nutrition knowledge questionnaire: an aid to assess the nutrition friendliness of schools of Dehradun, Uttarakhand EKTA BELWAL, ARCHANA KUSHWAHA, SARITA SRIVASTAVA, C.S. CHOPRA and ANIL KUMAR SHUKLA	452
Potential of common leaves of India as a source of Leaf Protein Concentrate RUSHDA ANAM MALIK, SHAYANI BOSE, ANURADHA DUTTA, DEEPA JOSHI, NIVEDITA, N.C. SHAHI, RAMAN MANOHARLALand G.V.S. SAIPRASAD	460
Job strain and muscle fatigue in small scale unorganized agri enterprises DEEPA VINAY, SEEMA KWATRA, SUNEETA SHARMA and KANCHAN SHILLA	466
Drudgery reduction of farm women involved in weeding of soybean crop SHALINI CHAKRABORTY	475
Childhood obesity and its association with hypertension among school-going children of Dehradun, Uttarakhand EKTA BELWAL, K. UMA DEVI and APARNA KUNA	482
Spring water and it's quality assessment for drinking purpose: A review SURABHI CHAND, H.J. PRASAD and JYOTHI PRASAD	489
Spatial distribution of water quality for Indo-Gangetic alluvial plain using Q-GIS SONALI KUMARA, VINOD KUMAR and ARVIND SINGH TOMAR	497
Application of geospatial techniques in morphometric analysis of sub-watersheds of Nanak Sagar Catchment AISHWARYA AWARI, DHEERAJ KUMAR, PANKAJ KUMAR, R. P. SINGH and YOGENDRA KUMAR	505
Evaluation of selected carbon sources in biofloc production and carps growth performance HAZIQ QAYOOM LONE, ASHUTOSH MISHRA, HEMA TEWARI, R.N. RAM and N.N. PANDEY	516
Calcium phosphate nanoparticles: a potential vaccine adjuvant YASHPAL SINGH and MUMTESH KUMAR SAXENA	523
Factors affecting some economic traits in Sahiwal Cattle DEVESH SINGH, C. B. SINGH, SHIVE KUMAR, B.N. SHAHI, BALVIR SINGH KHADDA, S. B. BHARDWAJ and SHIWANSHU TIWARI	528
The effect of probiotics and growth stimulants on growth performance of Murrah Buffalo SAMEER PANDEY, RAJ KUMAR, D.S. SAHU, SHIWANSHU TIWARI, PAWAN KUMAR, ATUL SHARMAand KARTIK TOMAR	532

Calcium phosphate nanoparticles: a potential vaccine adjuvant

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ABSTRACT: Nanotechnology is a branch of science and engineering, which plays a crucial role in numerous areas like medical, agricultural, pharmaceutical sector. Various nanoparticles are considered to be efficient drug delivery carrier. The usage of nanoparticles in vaccine formulation facilitates improved antigen stability, immunogenicity as well as slow release and targeted delivery. Several organic and inorganic nanoparticles have been evaluated for application in nanomedicines for targeted drug delivery as vaccine adjuvant system. Various Polymeric nanoparticles such as Poly Lactic Acid (PLA), Poly Lactic-Co-Glycolic Acid (PLGA), Polycaprolactone (PCL), Chitosan, Gelatin etc. have been studied as drug delivery agent for the treatment of diseases. Among inorganic nanoparticles, Calcium phosphate nanoparticles have been considered as most suitable and safe vaccine adjuvants due to their high biodegradability and biocompatibility. These nanoparticles are cost-effective and have hydrophilic nature, better stability and potential efficacy to produce better immune response in comparison to other nanoparticle adjuvant system.

Key words: Calcium phosphate nanoparticles, Nanotechnology, vaccine adjuvants

Adjuvants can be defined as substance or material, which comprises of chemically complex molecules that enhances an immune response to an antigen (Schijns and Lavelle, 2011). They have been used to enhance and improve the efficacy of vaccines from last century. The adjuvants act by different ways such as by depot generation, improving antigen presentation or by immunomodulation (Reed et al., 2013). Several particulate and non-particulate adjuvants have been used to enhance the immunity response to an antigen. Particulate adjuvants such as mineral salts (alum), lipid particulate and microparticles are most commonly used in human and animal vaccines while non-particulate adjuvants (Saponins, mucosal adjuvants, Chitosan etc.) also play an important role in enhancing the immunogenicity. Advancement in the field of Nanotechnology offers the opportunity to design nanoparticles varying in size, shape, composition and surface properties, for application in the field of medicine (Couvreur and Vauthier, 2006; Moghimi et al., 2005). These nano-sized particles are able to enter living cells and play a crucial role in diagnosis of diseases as well as drug delivery for exploring the treatment and prevention of various diseases such as Alzheimer's (Chackerian, 2010), hypertension (Tissot et al., 2008) and nicotine addiction (Maurer

et al., 2005). Several nanoparticles such as Gold nanoparticles, Carbon nanoparticles, Silver nanoparticles, mesoporous silica nanoparticles etc. are most commonly used as drug delivery agent as well as diagnostic agents (Chen *et al.*, 2012; Wang *et al.*, 2011; Yu *et al.*, 2013).

Calcium phosphate nanoparticles as nanoadjuvant system

Nanotechnology plays a pivotal role in development of new generation and effective adjuvant system. The application of nanoparticles in vaccines enhanced targeted delivery, immunogenicity and stability. In past few years, nanoparticles of varying size and composition have been used as adjuvant to enhance immune potential of antigens and as drug delivery system for prevention and treatment of disease (Couvreur and Vauthier, 2006). Several different studies have reported the application of nano-materials as adjuvant for human use and many of them are still under clinical trials (Correia-Pinto et al., 2013). The nano-based adjuvant system is divided into organic and inorganic nanoparticles. The organic nanoparticles include polymeric nanoparticles such as Poly (d,l-lactide-co-glycolide) (PLG), poly(ethylene glycol) (PEG) (Vila et al., 2004) poly(g-glutamic acid) (g-PGA) (Akagi et al.,

in vivo(Chu et al., 2017). Initially, it was used as an adjuvant of human vaccine (Diptheria-tetanuspertussis) in France (Goto et al., 1997; Masson et al., 2017). Calcium phosphate nanoparticles are made up of Calcium, Phosphorous and Oxygen therefore these have high biocompatibility

Synthesis of calcium phosphate nanoparticles

biodegradability in comparison to other

nanoparticles likes Zn and Cu (Lalk et al., 2013).

The Calcium phosphate nanoparticles are generally synthesized by following two approaches i.e,. Top down approach, which involves breakage of large sized microparticles into nano-sized particles by ultrasonication method (Bisht et al., 2005) and Top up approach that involves Co-precipitation method for synthesis of Calcium phosphate nanoparticles(He et al., 2002; Joyappa et al., 2009). Calcium phosphate nanoparticles synthesized by Co-precipitation method were used as adjuvant for vaccine development and provoked an effective immune response by Depot effect (Jones et al., 2014) and NLRP-3 inflammasome activation (Ronchi et al., 2016) as well as protective immunity against diseases like New castle disease, Human enterovirus-71, Friend rotavirus (Koppad et al., 2011; Saeed et al., 2015; Knuschke et al., 2016). These nanoparticles are more stable in vivo in comparison to polymeric nanoparticles (as polyacrylic acid nanoparticles) (Colby et al., 2013). In addition, the sterility and stability of Calcium phosphate nanoparticles In vitro are easier to control due to inorganic nature of Calcium phosphate nanoparticles. These nanoparticles are very costeffective in comparison to other nanoparticles (Castro et al., 2016).

Mechanism of action

Calcium phosphate nanoparticles produce immune response by Depot effect (Jones et al., 2014), NLRP-3 inflammasome activation (Ronchi et al., 2016) and produce a balanced cell mediated and humoral immune response by activation of Th1 and Th2 responses. The small size of nanoparticles facilitates the uptake by macrophages. Therefore, Calcium phosphate nanoparticles have been used as adjuvant for vaccines of animals (FMD, New castle disease,

2011) and poly(d,l-lactic-coglycolic acid)(PLGA) (Lu et al., 2009) which have been evaluated as vaccine and drug delivery systems due to biodegradability and efficient biocompatibility. Apart from synthetic polymeric nanoparticles, natural polymers such as chitosan (Feng et al., 2013), inulin (Saade et al., 2013) and alginate (Li et al., 2013) were also used as adjuvant system for delivery of antigens. These natural polymers have high biocompatibility, high biodegradability (Chua et al., 2012; Akagi et al., 2011). Therefore, they were used for animal (New castle disease) and human (HBV) vaccines (Borges et al., 2008; Zhao et al., 2012). On the other hand inorganic nanoparticles have better stability and resistance from microbial attack. They can be prepared at low temperature and are relatively inexpensive than organic nanoparticles. Inorganic nanoparticles can be further divided into two categories i.e., metallic and non-metallic nanoparticles. Metallic nanoparticles like gold and silver nanoparticles have been used for delivery of antigens for various diseases like FMD (Chen et al., 2010), influenza (Tao et al., 2014) human immunodeficiency virus-acquired immunity deficiency syndrome (HIV-AIDS) (Xu et al., 2012) and generated an effective immune response but the toxicity caused by metallic nanoparticles is a debatable issue (Lin et al., 2017). Non-metallic nanoparticles such as Silica nanoparticles, Carbon nanoparticles and Calcium phosphate nanoparticles were developed for drug delivery and adjuvant system (He et al., 2002;Yu et al., 2013; He et al., 2000). Among these non-metallic inorganic nanoparticles, Calcium phosphate nanoparticles have immense potential to function as nano adjuvants for inducing more balanced T helper type (Th1 and Th2) immune responses for better cell mediated and humoral immunity (Lin et al., 2017). These nanoparticles are highly biocompatible, biodegradable and non-toxic for the cells (Joyappa et al., 2009). These nanoparticles were mainly used for delivery of DNA vaccines (He et al., 2002;He et al., 2000; Joyappa et al., 2009).Calcium phosphate nanoparticles have been extensively used in therapeutics and diagnostics (Desai and Uskokovic, 2013). Calcium phosphate is a normal constituent of body that is well tolerated and rapidly absorbed

and

Aeromonas vaccines) and humans (Human enterovirus-71, Herpes simplex virus type2 and Toxoplasma gondii) (Doel, 2003; Zhang *et al.*, 2013;Saeed *et al.*, 2015).

Advantages of Calcium phosphate nanoparticles Calcium phosphate nanoparticles have a promising potential as targeted drug delivery agents. These nanoparticles are easy to deliver, cost-effective and readily absorbed, which allows their extensive use for entrapment of biomolecules. Various surface modifiers such as Cellobiose, Alginate, Gelatin etc. can be used for surface modification which leads to increased loading and releasing capacity of the drug in nanoparticles. The nano size of these nanoparticles helps in immobilizing and targeting biomolecules at specific site as they have high penetrability acting directly at cellular level. Several properties such as particle size, surface modification, surface charge etc. influence the potential of Calcium phosphate nanoparticles as drug delivery agent as well as efficient nano delivery adjuvant system for antigens.

CONCLUSION AND FUTURE PERSPECTIVES

Calcium phosphate nanoparticles have been proven suitable and safe for their use as a nano delivery adjuvant system in human vaccines. These nanoparticles are highly biocompatible and biodegradable and considered to be non-toxic at cellular level. Calcium phosphate nanoparticles have been reported to generate an effective cellular as well as humoral immune response and also can play a crucial role in site specific drug delivery system for many diseases at cellular level.

REFERENCES

- Akagi, T., Baba, M. and Akashi, M. (2011). Biodegradable nanoparticles as vaccine adjuvants and delivery systems: regulation of immune responses by nanoparticle-based vaccine. In *Polymers in nanomedicine*, Springer, Berlin, Heidelberg, Pp. 31-64.
- Bisht, S., Bhakta, G., Mitra, S. and Maitra, A. (2005). pDNA loaded calcium phosphate

nanoparticles: highly efficient non-viral vector for gene delivery. *International Journal of Pharmaceutics*, 288(1): 157-168

- Borges, O., Cordeiro-da-Silva, A., Tavares, J., Santarém, N., de Sousa, A., Borchard, G and Junginger, H.E. (2008). Immune response by nasal delivery of hepatitis B surface antigen and co-delivery of a CpG ODN in alginate coated chitosan nanoparticles. *European Journal of Pharmaceutics and Biopharmaceutics*, 69(2): 405-416.
- Castro, F., Ribeiro, V.P., Ferreira, A. (2016). Continuous flow precipitation as a route to prepare highly controlled nano hydroxyapatite *In vitro* mineralization and biological evaluation. *Matter Res Expression*, 3(7):075404.
- Chen, K., Zhang, J. and Gu, H. (2012). Dissolution from inside: a unique degradation behaviour of core-shell magnetic mesoporous silica nanoparticles and the effect of polyethyleneimine coating. *Journal of Materials Chemistry*, 22(41): 22005-22012.
- Chen, Y.S., Hung, Y.C., Lin, W.H. and Huang, G.S. (2010). Assessment of gold nanoparticles as a size-dependent vaccine carrier for enhancing the antibody response against synthetic foot-and-mouth disease virus peptide. *Nanotechnology*, 21(19): 195101.
- Chua, B.Y., Al Kobaisi, M., Zeng, W., Mainwaring, D. and Jackson, D.C. (2012). Chitosan microparticles and nanoparticles as biocompatible delivery vehicles for peptide and protein-based immune contraceptive vaccines. *Molecular Pharmaceutics*, 9(1): 81-90.
- Chu, W., Huang, Y., Yang, C., Liao, Y., Zhang, X., Yan, M., Cui, S. and Zhao, C. (2017). Calcium phosphate nanoparticles functionalized with alendronate-conjugated polyethylene glycol (PEG) for the treatment of bone metastasis. *International Journal of Pharmaceutics*, 516(1-2): 352-363.
- Chackerian, B. (2010). Virus-like particlebased vaccines for Alzheimer disease. *Human Vaccines*, 6:926–30.
- Couvreur, P. and Vauthier, C. (2006).

Nanotechnology: intelligent design to treat complex disease. *Pharmaceutical Research*, 23(7): 1417-1450.

- Correia-Pinto, J.F., Csaba, N. and Alonso, M.J. (2013). Vaccine delivery carriers: insights and future perspectives. *International Journal of Pharmaceutics*, 440(1): 27-38.
- Colby, A.H., Colson, Y.L., Grinstaff, M.W. (2013). Microscopy and tunable resistive pulse sensing characterization of the swelling of pH-responsive, polymeric expansile nanoparticles. *Nanoscale*, 5(8): 3496-3504.
- Desai, T.A. and Uskoković, V. (2013). Calcium phosphate nanoparticles: a future therapeutic platform for the treatment of osteomyelitis?. *Therapeutic Delivery*, 4(6): 643-645.
- Doel, T.R. (2003). FMD Vaccines. *Virus Res*, 91(1): 81-99.
- Feng, G., Jiang, Q., Xia, M., Lu, Y., Qiu, W., Zhao,
 D., Lu, L., Peng, G. and Wang, Y. (2013).
 Enhanced immune response and protective effects of nano-chitosan-based DNA vaccine encoding T cell epitopes of Esat-6 and FL against *Mycobacterium tuberculosis* infection. *PLoS One*, 8(4): pe61135
- Goto, N., Kato, H., Maeyama, J.I., Shibano, M., Saito, T., Yamaguchi, J. and Yoshihara, S. (1997). Local tissue irritating effects and adjuvant activities of calcium phosphate and aluminium hydroxide with different physical properties. *Vaccine*, 15(12-13): 1364-1371.
- He, Q., Mitchell, A., Morcol, T. and Bell, S.J. (2002). Calcium phosphate nanoparticles induce mucosal immunity and protection against herpes simplex virus type 2. *Clinical and Diagnostic Laboratory Immunology*, 9(5): 1021-1024.
- He, Q., Mitchell, A.R., Johnson, S.L., Wagner-Bartak, C., Morcol, T. and Bell, S.J. (2000). Calcium phosphate nanoparticle adjuvant. *Clinical and Diagnostic Laboratory Immunology*, 7(6): 899-903.
- Joyappa, D.H., Kumar, C.A., Banumathi, N., Reddy, G.R. and Suryanarayana, V.V. (2009). Calcium phosphate nanoparticle prepared

with foot and mouth disease virus P1-3CD gene construct protects mice and guinea pigs against the challenge virus. *Veterinary Microbiology*, 139(1-2): 58-66.

- Jones, S., Asokanathan, C. and Kmiec, D. (2014). Protein coated micro crystals formulated with model antigens and modified with calcium phosphate exhibit enhanced phagocytosis and immunogenicity. *Vaccine*, 32(33): 4234-4342.
- Knuschke, T., Rotan, O., Bayer, W., Sokolova, V., Hansen, W., Sparwasser, T., Dittmer, U., Epple, M., Buer, J. and Westendorf, A.M. (2016). Combination of nanoparticle-based therapeutic vaccination and transient ablation of regulatory T cells enhances antiviral immunity during chronic retroviral infection. *Retrovirology*, 13(1): 24-34.
- Koppad, S., Raj, G.D., Gopinath, V.P., Kirubaharan, J.J., Thangavelu, A. and Thiagarajan, V. (2011). Calcium phosphate coupled Newcastle disease vaccine elicits humoral and cell mediated immune responses in chickens. *Research in Veterinary Science*, 91(3): 384-390.
- Lalk, M., Reifenrath, J., Angrisani, N., Bondarenko, A., Seitz, J.M., Mueller, P.P. and Meyer-Lindenberg, A. (2013). Fluoride and calcium-phosphate coated sponges of the magnesium alloy AX30 as bone grafts: a comparative study in rabbits. *Journal of Materials Science: Materials in Medicine*, 24(2): 417-436.
- Li, P., Luo, Z., Liu, P., Gao, N., Zhang, Y., Pan, H., Liu, L., Wang, C., Cai, L. and Ma, Y. (2013). Bio reducible alginate-poly (ethylenimine) nanogels as an antigen-delivery system robustly enhance vaccine-elicited humoral and cellular immune responses. *Journal of Controlled Release*, 168(3): 271-279.
- Lin, Y., Wang, X., Huang, X., Zhang, J., Xia, N. and Zhao, Q. (2017). Calcium phosphate nanoparticles as a new generation vaccine adjuvant. *Expert review of vaccines*, 16(9): 895-906.
- Lu, J.M., Wang, X., Marin-Muller, C., Wang, H., Lin, P.H., Yao, Q. and Chen, C. (2009). Current

advances in research and clinical applications of PLGA-based nanotechnology. *Expert review of Molecular Diagnostics*, 9(4): 325-341.

- Masson, J.D., Thibaudon, M., Bélec, L. and Crépeaux, G. (2017). Calcium phosphate: a substitute for aluminum adjuvants? *Expert Review of Vaccines*, 16(3): 289-299.
- Maurer, P., Jennings, G.T., Willers, J., Rohner, F., Lindman, Y. and Roubicek K. (2005).A therapeutic vaccine for nicotine dependence: preclinical efficacy, and phase I safety and immunogenicity. *European Journal of Immunology*, 35: 2031–40.
- Moghimi, S.M., Hunter, A.C. and Murray, J.C. (2005). Nanomedicine: current status and future prospects. *The FASEB Journal*, 19:311–30.
- Reed, S.G., Orr, M.T. and Fox, C.B. (2013). Key roles of adjuvants in modern vaccines. *Nature Medicine*, 19(12): 1597-1608.
- Saade, F., Honda-Okubo, Y., Trec, S. and Petrovsky, N. (2013). A novel hepatitis B vaccine containing AdvaxTM, a polysaccharide adjuvant derived from delta inulin, induces robust humoral and cellular immunity with minimal reactogenicity in preclinical testing. *Vaccine*, 31(15):1999-2007.
- Saeed, M.I., Omar, A.R., Hussein, M.Z., Elkhidir, I.M. and Sekawi, Z. (2015). Systemic antibody response to nano-size calcium phospate biocompatible adjuvant adsorbed HEV-71 killed vaccine. *Clinical and Experimental Vaccine Research*, 4(1): .88-98.
- Schijns, V. E. and Lavelle, E.C. (2011). Trends in vaccine adjuvants. *Expert review of vaccines*, 10(4): 539-550.
- Ronchi, F., Basso, C and Preite, S. (2016). Experimental priming of encephalitogenic Th1/Th17 cells requires pertussis toxindriven IL-1β production by myeloid cells. *Nature Communication*, 7:11541.
- Tao, W., Ziemer, K.S. and Gill, H.S. (2014). Gold nanoparticle–M2e conjugate co-formulated with CpG induces protective immunity

against influenza A virus. *Nanomedicine*, 9(2): 237-251.

- Tissot, A.C., Maurer, P., Nussberger, J., Sabat, R., Pfister, T. and Ignatenko, S. (2008). Effect of immunization against angiotensin II with CYT006-AngQb on ambulatory blood pressure: a double-blind, randomized, placebo-controlled phase II a study. *The Lancet*, 371:821–7.
- Vila, A., Sanchez, A., Evora, C., Soriano, I., Vila Jato, J.L. and Alonso, M.J. (2004). PEG-PLA nanoparticles as carriers for nasal vaccine delivery. *Journal of Aerosol* Medicine, 17(2): 174-185.
- Wang, T., Zou, M., Jiang, H., Ji, Z., Gao, P. and Cheng, G. (2011). Synthesis of a novel kind of carbon nanoparticle with large mesopores and macropores and its application as an oral vaccine adjuvant. *European Journal of Pharmaceutical Sciences*, 44(5): 653-659.
- Xu, L., Liu, Y., Chen, Z., Li, W., Liu, Y., Wang, L., Liu, Y., Wu, X., Ji, Y., Zhao, Y. and Ma, L. (2012). Surface-engineered gold nanorods: promising DNA vaccine adjuvant for HIV-1 treatment. *Nano Letters*, 12(4): 2003-2012.
- Yu, M., Jambhrunkar, S., Thorn, P., Chen, J., Gu, W. and Yu, C. (2013). Hyaluronic acid modified mesoporous silica nanoparticles for targeted drug delivery to CD44overexpressing cancer cells. *Nanoscale*, 5(1): 178-183.
- Zhang, N.Z., Chen, J. and Wang, M. (2013). Vaccine against Toxoplasma gondii: new developments and perspectives. *Expert Review of Vaccines*, 12(11): 1287-1299.
- Zhao, K., Chen, G., Shi, X.M., Gao, T.T., Li, W., Zhao, Y., Zhang, F.Q., Wu, J., Cui, X. and Wang, Y.F. (2012). Preparation and efficacy of a live newcastle disease virus vaccine encapsulated in chitosan nanoparticles. *PLoS One*, 7(12): 53314.

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