Print ISSN: 0972-8813 e-ISSN: 2582-2780 [Vol. 22(1) January-April 2024]

Pantnagar Journal of Research

(Formerly International Journal of Basic and Applied Agricultural Research ISSN: 2349-8765)



G.B. Pant University of Agriculture & Technology, Pantnagar

ADVISORYBOARD

Patron

Dr. Manmohan Singh Chauhan, Vice-Chancellor, G.B. Pant University of Agriculture and Technology, Pantnagar, India Members

Dr. A.S. Nain, Ph.D., Director Research, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. Jitendra Kwatra, Ph.D., Director, Extension Education, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. S.K. Kashyap, Ph.D., Dean, College of Agriculture, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. S.P. Singh, Ph.D., Dean, College of Veterinary & Animal Sciences, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. K.P. Raverkar, Ph.D., Dean, College of Post Graduate Studies, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. Sandeep Arora, Ph.D., Dean, College of Basic Sciences & Humanities, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. Alaknanda Ashok, Ph.D., Dean, College of Technology, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. Alka Goel, Ph.D., Dean, College of Community Science, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. Aydhesh Kumar, Ph.D., Dean, College of Fisheries, G.B. Pant University of Agri. & Tech., Pantnagar, India

Dr. R.S. Jadoun, Ph.D., Dean, College of Agribusiness Management, G.B. Pant University of Agri. & Tech., Pantnagar, India

EDITORIALBOARD

Members

Prof. A.K. Misra, Ph.D., Chairman, Agricultural Scientists Recruitment Board, Krishi Anusandhan Bhavan I, New Delhi, India Dr. Anand Shukla, Director, Reefberry Foodex Pvt. Ltd., Veraval, Gujarat, India

Dr. Anil Kumar, Ph.D., Director, Education, Rani Lakshmi Bai Central Agricultural University, Jhansi, India

Dr. Ashok K. Mishra, Ph.D., Kemper and Ethel Marley Foundation Chair, W.P. Carey Business School, Arizona State University, U.S.A.

Dr. B.B. Singh, Ph.D., Visiting Professor and Senior Fellow, Dept. of Soil and Crop Sciences and Borlaug Institute for International Agriculture, Texas A&M University, U.S.A.

Prof. Binod Kumar Kanaujia, Ph.D., Professor, School of Computational and Integrative Sciences, Jawahar Lal Nehru University, New Delhi, India

Dr. D. Ratna Kumari, Ph.D., Associate Dean, College of Community / Home Science, PJTSAU, Hyderabad, India

Dr. Deepak Pant, Ph.D., Separation and Conversion Technology, Flemish Institute for Technological Research (VITO), Belgium

Dr. Desirazu N. Rao, Ph.D., Professor, Department of Biochemistry, Indian Institute of Science, Bangalore, India

Dr. G.K. Garg, Ph.D., Dean (Retired), College of Basic Sciences & Humanities, G.B. Pant University of Agric. & Tech., Pantnagar, India

Dr. Humnath Bhandari, Ph.D., IRRI Representative for Bangladesh, Agricultural Economist, Agrifood Policy Platform, Philippines

Dr. Indu S Sawant, Ph.D., Director, ICAR - National Research Centre for Grapes, Pune, India

Dr. Kuldeep Singh, Ph.D., Director, ICAR - National Bureau of Plant Genetic Resources, New Delhi, India

Dr. M.P. Pandey, Ph.D., Ex. Vice Chancellor, BAU, Ranchi & IGKV, Raipur and Director General, IAT, Allahabad, India

Dr. Martin Mortimer, Ph.D., Professor, The Centre of Excellence for Sustainable Food Systems, University of Liverpool, United Kingdom

Dr. Muneshwar Singh, Ph.D., Project Coordinator AICRP-LTFE, ICAR - Indian Institute of Soil Science, Bhopal, India

 $Prof.\ Omkar, Ph.D., Professor, Department\ of\ Zoology, University\ of\ Lucknow, India$

Dr. P.C. Srivastav, Ph.D., Professor, Department of Soil Science, G.B. Pant University of Agriculture and Technology, Pantnagar, India

Dr. Prashant Srivastava, Ph.D., Cooperative Research Centre for Contamination Assessment and Remediation of the Environment, University of South Australia. Australia

Dr. Puneet Srivastava, Ph.D., Director, Water Resources Center, Butler-Cunningham Eminent Scholar, Professor, Biosystems Engineering, Auburn University, U.S.A.

Dr. R.C. Chaudhary, Ph.D., Chairman, Participatory Rural Development Foundation, Gorakhpur, India

Dr. R.K. Singh, Ph.D., Director & Vice Chancellor, ICAR-Indian Veterinary Research Institute, Izatnagar, U.P., India

Prof. Ramesh Kanwar, Ph.D., Charles F. Curtiss Distinguished Professor of Water Resources Engineering, Iowa State University, U.S.A.

Dr. S.N. Maurya, Ph.D., Professor (Retired), Department of Gynecology & Obstetrics, G.B. Pant University of Agric. & Tech., Pantnagar, India

Dr. Sham S. Goyal, Ph.D., Professor (Retired), Faculty of Agriculture and Environmental Sciences, University of California, Davis, U.S.A.

Prof. Umesh Varshney, Ph.D., Professor, Department of Microbiology and Cell Biology, Indian Institute of Science, Bangalore, India

Prof. V.D. Sharma, Ph.D., Dean Academics, SAI Group of Institutions, Dehradun, India

Dr. V.K. Singh, Ph.D., Head, Division of Agronomy, ICAR-Indian Agricultural Research Institute, New Delhi, India

Dr. Vijay P. Singh, Ph.D., Distinguished Professor, Caroline and William N. Lehrer Distinguished Chair in Water Engineering, Department of Biological Agricultural Engineering, Texas A&M University, U.S.A.

Dr. Vinay Mehrotra, Ph.D., President, Vinlax Canada Inc., Canada

Editor-in-Chief

Dr. Manoranjan Dutta, Head Crop Improvement Division (Retd.), National Bureau of Plant Genetic Resources, New Delhi, India

Managing Editor

 $Dr.\,S.N.\,Tiwari, Ph.D., Professor, Department of Entomology, G.B.\,Pant\,University\, of Agriculture\, and\,Technology, Pantnagar, India and Technology, Pantnagar, India and India$

Assistant Managing Editor

Dr. Jyotsna Yadav, Ph.D., Research Editor, Directorate of Research, G.B. Pant University of Agriculture and Technology, Pantnagar, India

Technical Manager

Dr. S.D. Samantray, Ph.D., Professor, Department of Computer Science and Engineering, G.B. Pant University of Agriculture and Technology, Pantnagar, India

PANTNAGAR JOURNAL OF RESEARCH

Vol. 22(1) January-April 2024

CONTENTS

| Productivity, nutrient uptake and economics of sweet corn (<i>Zea mays</i> L. var. <i>saccharata</i>) under different planting geometry and NPK levels AMIT BHATNAGAR, SAILESH DEB KARJEE, GURVINDER SINGH and DINESH KUMAR SINGH | 1-7 |
|---|--------|
| Integrated effect of natural farming concoctions and organic farming practices with various NPK doses on quality of bread wheat PRERNA NEGI, MOINUDDIN CHISTI and HIMANSHU VERMA | 8-13 |
| Characterization and fertility capability classification of some soils in the rain forest zone of Edo state, Nigeria OKUNSEBOR, F.E., OGBEMUDIA, I. and OKOLIE, S. I. | 14-25 |
| Characterization and classification of guava growing soils of North-East Haryana according to frame work of land evaluation (FAO, 1993) DHARAM PAL, MANOJ SHARMA, R.S. GARHWAL and DINESH | 26-35 |
| Interactive impact of heavy metals and mycorrhizal fungi on growth and yield of pepper (Capsicum annuum Linn.) SHARMILA CHAUHAN, MOHINDER SINGH, SNEHA DOBHAL, DEEKSHA SEMWAL and PRAVEEN | 36-47 |
| Response of chilli (<i>Capsicum annuum</i> var. <i>annuum</i> L.) to different nutrient management practices SHEETAL, K.C. SHARMA, SHIVAM SHARMA, NEHA SHARMA, D.R. CHAUDHARY, SANDEEP MANUJA and AKHILESH SHARMA | 48-58 |
| Trend detection in weather parameters using Mann-Kendall test for <i>Tarai</i> region of Uttarakhand SHUBHIKA GOEL and R.K. SINGH | 59-67 |
| Comparative study of antioxidant potential of fresh peel from different citrus species TARU NEGI, ANIL KUMAR, ARCHANA GANGWAR, SATISH KUMAR SHARMA, ANURADHA DUTTA, NAVIN CHAND SHAHI, OM PRAKASH and ASHUTOSH DUBEY | 68-74 |
| Suitability of Quinoa Grains (<i>Chenopodium Quinoa Willd.</i>) for development of Low Glycemic Index Biscuits RUSHDA ANAM MALIK, SARITA SRIVASTAVA and MEENAL | 75-84 |
| A study on dietary intake among school-going adolescent girls of Udaipur, Rajasthan during COVID-19 JYOTI SINGH and NIKITA WADHAWAN | 85-92 |
| Nutritional and sensory evaluation of gluten free chapatti developed using underutilised food sources AYUSHI JOSHI, ARCHANA KUSHWAHA, ANURADHA DUTTA, ANIL KUMAR and NAVIN CHANDRA SHAHI | 93-98 |
| Nutrient-enriched wheat <i>chapatti</i> with fresh pea shells (<i>Pisum sativum l.</i>): A comprehensive quality assessment AMITA BENIWAL, SAVITA, VEENU SANGWAN and DARSHAN PUNIA | 99-109 |

| Pearl Millet-Based Pasta and Noodles Incorporated with <i>Jamun</i> Seed Powder: Quality Analysis SAVITA, AMITA BENIWAL, VEENU SANGWAN and ASHA KAWATRA | 110-121 |
|---|---------|
| Unlocking the biofortification potential of <i>Serratia marcescens</i> for enhanced zinc and iron content in wheat grains BHARTI KUKRETI and AJAY VEER SINGH | 122-131 |
| Antioxidant and anti-inflammatory properties of sun-dried leaves and fruits of wild <i>Pyracantha crenulata</i> (D. Don) M. Roem. SUGANDHA PANT, PREETI CHATURVEDI, AAKANSHA VERMA, MANDEEP RAWAT, VAISHNAVI RAJWAR and KAVITA NEGI | 132-141 |
| Studies on productive herd life, longevity, and selective value and their components in crossbred cattle SHASHIKANT, C.V. SINGH and R.S. BARWAL | 142-150 |
| Studies on replacement rate and its components in crossbred cattle SHASHIKANT, C.V. SINGH, R.S. BARWAL and MANITA DANGI | 151-157 |
| Principal component analysis in production and reproduction traits of Frieswal cattle under field progeny testing OLYMPICA SARMA, R. S. BARWAL, C. V. SINGH, D. KUMAR, C. B. SINGH, A. K. GHOSH, B. N. SHAHI and S. K. SINGH | 158-163 |
| Degenerative renal pathology in swine: A comprehensive histopathological investigation in Rajasthan, India SHOBHA BURDAK, INDU VYAS, HEMANT DADHICH, MANISHA MATHUR, SHESH ASOPA, RENU | 164-169 |
| Evaluation of histopathological changes on acute exposure of profenofos in Swiss albino mice SONU DEVI, VINOD KUMAR, PREETI BAGRI and DEEPIKA LATHER | 170-177 |
| Temporal and spatial performance of rapeseed and mustard oilseed in India: A study in the context of Technology Mission on Oilseeds! LEKHA KALRA and S. K. SRIVASTAVA | 178-190 |
| Comparative economics of maize cultivation in major and minor maize producing districts of Karnataka – a study across farm size groups GEETHA, R. S. and S. K. SRIVASTAVA | 191-203 |
| A study on Usefulness of Participatory Newsletter for Potato growers in Udham Singh Nagar district of Uttarakhand RAMESH NAUTIYAL and ARPITA SHARMA KANDPAL | 204209 |
| Training Needs of Hortipreneurs in Value Addition and fruit crop production in Kumaon Hills of Uttarakhand KRITIKA PANT and ARPITA SHARMA KANDPAL | 210-215 |
| Post-training Knowledge Assessment of the rural women about Mushroom Cultivation under TSP project, funded by ICAR ARPITA SHARMA KANDPAL, S. K. MISHRAand OMVEER SINGH | 216-220 |
| UAV Technology: Applications, economical reliance and feasibility in Indian Agriculture A. AJAY and S. SAI MOHAN | 221-229 |

Comparative study of antioxidant potential of fresh peel from different citrus species

TARU NEGI¹, ANIL KUMAR¹*, ARCHANA GANGWAR¹, SATISH KUMAR SHARMA¹, ANURADHA DUTTA², NAVIN CHAND SHAHI³, OM PRAKASH⁴ and ASHUTOSH DUBEY⁵

¹Department of Food Science and Technology, College of Agriculture, ²Department of Foods and Nutrition, College of Community Sciences, ³Department of PHPFE, College of Technology, ⁴Department of Chemistry, ⁵Department of Biochemistry, College of Basic Sciences, G.B. Pant University of Agriculture and Technology, Pantnagar-263145(U.S. Nagar, Uttarakhand)

*Corresponding author's email id: anilkumargbpuat@gmail.com

ABSTRACT: Citrus peels contain high concentration of bioactive compounds, particularly phenols and flavonoids that have a positive impact on human health. In this study, phenol, flavonoid, and antioxidant potency composite index (APC Index) of fresh citrus peels (kinnow, orange, mosambi, hill lemon, and malta) were assessed. DPPH, FRAP, and ABTS assays were done to determine the APC Index of the peels. The study findings showed that, among fresh citrus peels, orange flavedo was found to contain the highest total phenols (15.11 mg GAE/g dw) whereas, kinnow flavedo had highest flavonoid (5.86 mg QCE/g dw). Highest APC Index was found in kinnow followed by orange, malta, hill lemon and mosambi. Therefore, it can be concluded from the present study that kinnow fresh peel had highest antioxidant potential as compared to other citrus peels taken in this study. It was also noted that the fresh peels of various citrus species have enormous source of bioactive compounds and high antioxidant capacity which has tremendous potential to be utilized in functional foods and as nutraceuticals.

Key words: Antioxidant potency composite index (APC index), bioactive compounds, citrus peel, flavonoid, phenol

Bioactive compounds (phenols, flavonoids etc) are secondary metabolites that are produced by plants in smaller quantities together with primary metabolites (carbohydrate, lipid, protein) and protect the plants from adverse conditions like drought. These secondary metabolites are known for health beneficial properties such as antiviral, antioxidant, antimicrobial and anti-inflammatory whose intake has been encouraged with the purpose to decrease cardiovascular diseases, hypertension, diabetes, and weight gain (Borgonovi et al., 2022). Food industry wastes (such as pulp, peel, stem, and seeds) contain a higher amount of bioactive compounds than fruit juice. Fruit peels are one of the food industry byproducts, which are commonly discarded during the consumption or manufacturing stage. Particularly, citrus peel waste (CPW) has immense economic value as it contains abundant amount of bioactive compounds (Sharma et al., 2017). According to reports, around 75% of total production of citrus fruit is used as fresh, while the remaining 25% is used for the development of commercial products (Mahato et al., 2019). The annual citrus waste generated in India is 7.8 million tons (Khan *et al.*, 2021). Consequently, the amount of citrus waste produced by processing enterprises is estimated to be over 60 million tons per year around the world (Mahato *et al.*, 2019). After processing, 50-60% of the fruit parts is mainly citrus waste (peels, seeds, and membrane residue) and 30-34% citrus peel waste (CPW) is generated which is known as a treasure of bioactive compounds (Rafiq *et al.*, 2018).

As already stated, bioactive compounds present in citrus peel are well known to have various health beneficial properties. Moreover, researchers discovered that flavonoids particularly, hesperidin, hesperetin, naringenin and essential oil of citrus fruit plays important role in preventing the coronavirus disease (COVID-19) caused by severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2). Citrus flavonoids, one of the bioactive compounds, prevent the binding of SARS-CoV-2 with angiotensin-converting enzyme 2 which is a cell membrane protein receptor for SARS-CoV-2 present in the host cell and therefore, inhibit the virus

replication (Alberca *et al.*, 2020, Bellavite and Donzelli 2020 and Agrawal *et al.*, 2021).

Hence, this study was carried out to compare the antioxidant properties of bioactives compounds extracted from fresh flavedo (outer colored part) of different citrus fruit which may act as an active ingredient for various foods or pharmaceutical products intended for improving immunity in human population.

MATERIALS AND METHODS

Collection of materials

Citrus fruits of different types i.e., Orange (Citrus sinensis), Mosambi (Citrus limetta), Kinnow (Citrus reticulata), Hill Lemon (Citrus pseudolimon) and Malta (Citrus sinensis) were procured from the local market of Pantnagar, India and were brought to the Department of Food Science and Technology, College of Agriculture, Govind Ballabh Pant University of Agriculture and Technology, Pantnagar, India.

Preparation of extract

Extract of fresh citrus flavedo was prepared according to the method of Jin *et al.* (2022) with slight modification. Fresh citrus flavedo (1 g) of the different citrus fruits under consideration were weighed and crushed in a pestle mortar. Solvent (20 mL) of absolute ethanol was added in it and extraction by reflux method was performed at 80°C for 2 h, filtered using a Whatman No. 1 filter paper and stored at 4°C till further use.

Physiochemical parameters *Moisture content*

The moisture content was estimated by drying the weighed sample up to a constant weight in hot air oven at $105 \pm 2\dot{\text{UUC}}$ (Özcan *et al.*, 2021). The dried sample was then cooled to room temperature in desiccator prior to weighing. Moisture in terms of percentage was calculated as given below:

Moisture (%) =
$$\frac{\text{(Weight of fresh sample - Weight of dried sample)}}{\text{Weight of fresh sample}} \times 100$$

Total Phenolic Content

Folin–Ciocalteu method (Singleton *et al.*, 1999) was used for the estimation of total phenolic content (TPC) from citrus peel flavedo with slight modification. All experiments were performed in triplicates. A total of 300 μ L of prepared extract, 1700 μ L of distilled water and 100 μ L of Folin-Ciocalteu reagent (FCR) were vortexed. Following this, 300 μ L of sodium carbonate (20 %) was added and mixed thoroughly. The mixture was kept for 60 min in a dark place and the absorbance was measured at 750 nm in a UV-Vis spectrophotometer. TPC was expressed in mg of gallic acid equivalent per gram of peel (mg GAE/g dry weight basis, dw).

Total Flavonoid Content

Total flavonoid content (TFC) was estimated following the method of Formagio *et al.* (2014) with slight modifications. All experiments were performed in triplicates. Extract (80 µL) was taken in a test tube, 0.42 mL of 95 % ethanol was added followed by addition of 0.025 mL aluminum chloride (10 %), 0.025 mL sodium acetate solution (1 M) and 0.7 mL distilled water. Solution was mixed and incubated for 40 min and the absorbance was measured at 415 nm. TFC was expressed in mg quercetin equivalents per gram of peel (mg QCE/g dw).

Antioxidant activity assays and Antioxidant potency composite index (APCI)

Antioxidant activity was measured using three different in vitro assays i.e., 2, 2-Diphenyl-1-2,2-azinobis-3picryhydrazyl (DPPH), ethylbenzthiazoline-6-sulphonic acid (ABTS) and Ferric reducing antioxidant power (FRAP). Method for measuring antioxidant activity by DPPH method was adopted from Abu Bakar et al. (2009) with slight modifications. An aliquot of citrus flavedo extract (130 μL) mixed with 5 mL of DPPH (0.004 %) dissolved in methanol and incubated at room temperature for 30 min. The final absorbance was measured at 517 nm using the UV-visible spectrophotometer while methanol was taken as blank. ABTS in vitro antioxidant method was performed according to the method of Belwal et al. (2016). Absorbance was measured at 734 nm using the UV-visible Spectrophotometer. For FRAP, the method of Singanusong et al. (2014) was followed. Absorbance was taken at 593 nm with the help of UV-visible spectrophotometer after incubating the sample in dark at 25 ± 2 °C for 30 min. Results were expressed in mg of BHT equivalent per gram of citrus peel (mg BHTE/g dw).

Antioxidant Potency Composite Index (APCI) of DPPH, ABTS and FRAP were calculated according to the method of Chen et al. (2021) using the following equation:

 $APCI = \frac{\text{measured value of assay (sample score)}}{\text{x 100}}$ maximum value of assay (best score)

Further, the average of all APCI index (DPPH, ABTS, FRAP) was calculated for each fresh citrus flavedo which was considered as APCI.

Statistical analysis

Data was expressed as mean \pm standard deviation of three replicates. One - way ANOVA with posthoc Tukey's multiple comparison test was performed for phenol, flavonoid and antioxidant capacity analysis. The graphical representation of data was done using GraphPad Prism 8.0.1 version software.

RESULTS AND DISCUSSION

Moisture content

Fresh hill lemon peel contained high moisture content (80.76 ± 0.83 %) followed by kinnow (76.10 \pm 0.56 %), malta (74.23 \pm 0.23), orange (73.82 \pm 0.91) and mousambi peel (62.56 \pm 1.28). Like our study, Abd El-ghfar et al. (2016) also reported more moisture content of fresh lemon peel (81.23 %) as compared to orange peel (74.35 %). Özcan et al. (2021) noted 82.08 % of moisture content in fresh lemon peel which was similar to our findings.

Total Phenol Content and Total Flavonoid Content

Both phenol and flavonoid play a vital role in human health as they show antioxidant, antibacterial, antiinflammatory, and anti-cancerous properties and, protect the human skin from UV rays (Tungmunnithum et al., 2018). Orange showed the highest phenol content (15.11 \pm 0.18 mg GAE/g dw) followed by kinnow (14.57 \pm 0.41 mg GAE/g dw), malta $(12.03 \pm 0.99 \text{ mg GAE/g dw})$, mosambi (11.45) \pm 0.87 mg GAE/g dw) and hill lemon (9.80 \pm 0.24 mg GAE/g dw) (Fig. 1A). Statistically, total phenol content of orange and kinnow peel were found at par with each other. In case of TFC, kinnow showed

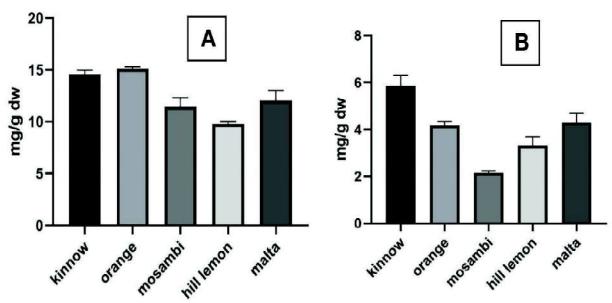


Fig. 1: (A) Total phenol content (mg GAE/g dw) (B) Total flavonoid content (mg QCE/g dw) in different fresh citrus flavedo, Error bars indicate the SD (n=3)

| Table 17 Inviolation were regional for the first control of the first co | | | | | | | | | | |
|--|----------------------|------------|--------------------------------|------------|-----------------------|------------|---------|---------|--|--|
| Citrus peel | DPPH | DPPH index | ABTS | ABTS index | FRAP | FRAP index | Average | Ranking | | |
| Kinnow | 60.41 ± 0.71 | 100.00 | 26.34 ± 1.32^{abc} | 100.00 | 38.59 ± 4.87^{ab} | 88.71 | 96.23 | 1 | | |
| Orange | 56.26 ± 0.45 | 93.13 | 22.71 ± 2.57^{adef} | 86.22 | 41.33 ± 5.82^{ac} | 95.01 | 91.45 | 2 | | |
| Mosambi | 45.29 ± 0.50 | 74.97 | 16.34 ± 2.25^{dgh} | 62.01 | 19.98 ± 1.47^{d} | 45.93 | 60.97 | 5 | | |
| Hill Lemon | 49.81 ± 0.41^a | 82.46 | $19.83 \pm 1.83^{\text{begi}}$ | 75.26 | 26.39 ± 3.56^d | 60.69 | 72.80 | 4 | | |
| Malta | 49.15 ± 1.03^{a} | 81 36 | $22.45 \pm 3.67^{\text{cfhi}}$ | 85.21 | 43.49 ± 2.48^{bc} | 100 | 88 85 | 3 | | |

Table 1: Antioxidant activity (mg/g dw) and APCI with their ranking in different citrus fruits

same lowercase letters show statistically non-significant difference within column, p < 0.05

highest TFC $(5.86 \pm 0.46 \text{ mg QCE/g dw})$ followed by malta $(4.29 \pm 0.99 \text{ mg QCE/g dw})$, orange $(4.19 \pm 0.16 \text{ mg QCE/g dw})$, hill lemon $(3.32 \pm 0.37 \text{ mg QCE/g dw})$ and mosambi $(2.15 \pm 0.10 \text{ mg QCE/g dw})$. Saleem *et al.* (2023) reported highest TPC (21.33 mg GAE/g) and TFC (2.02 mg QCE/g dw) in orange peel and lemon peel, respectively. Assefa *et al.* (2017) reported total flavonoid content of 2.1 mg/g g dw in orange peel which was slightly less than our study. Different harvesting time, cultivar, extraction method, extract drying and geographical region are some of the important factors which may be responsible for the variations in the content of phenol and flavonoid in different studies (Rafiq *et al.*, 2019).

Antioxidant activity and Antioxidant potency composite index (APCI)

Antioxidant assay generally works on two principal modes of action i.e., hydrogen atom transfer (HAT) and single electron transfer (SET). DPPH and ABTS follow HAT mechanism whereas FRAP works on SET mechanism (Norma Francenia et al., 2019, Siddeeg et al., 2021). In DPPH assay, deep purple colour of DPPH is changed to pale yellow colour (2,2-diphenyl-1-picrylhydrazine) by the antioxidants present in sample whereas in ABTS assay, ABTS blue-green radical is generated that is reduced by antioxidants and changes its colour into pale blue (Kim et al., 2023). On the contrary, FRAP assay examines the overall reducing ability of antioxidants present in the sample by estimating the reduction of ferric (III) ion to ferrous (II) ions (Li et al., 2008). Proper selection of methods is necessary for the determination of antioxidant capacity, based on the reaction characteristics of the compounds, as there is no single method for determining antioxidant capacity that reflects the effect of all antioxidants present in a complex mixture of bioactive substances due to the different mechanisms of action and nature of antioxidant and oxidizing substances (Gómez-Urios et al., 2023). Therefore, it is always preferable to check antioxidant activity of sample by more than one method. Table 1 shows the antioxidant capacity of citrus flavedo extract measured by different antioxidant assays i.e., DPPH, ABTS and FRAP assay. Kinnow flavedo showed the highest DPPH (60.41 mg/g dw) and ABTS (26.34 mg/g dw) antioxidant capacity as compared to other citrus flavedo which might be due to the presence of high phenol and flavonoid content. However, FRAP antioxidant capacity of orange flavedo (41.33 mg/g dw) was found at par with kinnow flavedo (38.59 mg/g dw). Chen et al. (2021) measured strong antioxidant capacity in mandarin and weakest in pummelo freeze-dried citrus peel powder extracted under ultrasonicated conditions with methanol.

All three assays i.e. DPPH, ABTS and FRAP gave different values of antioxidant capacities for citrus flavedo extract. Since a single method for antioxidant estimation may not be very reliable, APCI was determined to take into consideration the overall effect of almost all the active ingredients/ bioactive components contributing towards antioxidant activity. In this context, APCI took into consideration DPPH, ABTS and FRAP for depicting overall antioxidant potential of the samples. As shown in Table 1, the average for DPPH, ABTS and FRAP APCI (i.e. overall APCI) ranged between 60.97 to 96.23 for fresh citrus flavedo based on which different ranks were assigned to the samples. Considering this ranking system, fresh kinnow citrus flavedo was ranked first whereas mosambi ranked last, among all citrus flavedo peel. Chen et al. (2020) computed the APCI of 27 citrus cultivars peels of eight citrus species which ranged from 10.06 to 80.13 and the top six APC index was found under Citrus reticulata species. Kinnow mandarin that ranked first in our study, also comes under *Citrus reticulata* species. Likewise, Chen *et al.* (2021) calculated APCI of 52 varieties including mandarin (27 cultivars), sweet orange (15 cultivars), pummelo (7 cultivars), lemon (2 species) and kumquat (1 cultivar) varied between 6.23 to 94.56 and mandarin showed the highest antioxidant capacity among all.

CONCLUSION

The results of this study showed that fresh citrus flavedo contains high concentration of flavonoid and phenolic content. Orange peel was found to contain a higher quantity of phenol content whereas kinnow peel had high flavonoid content, among all citrus peels. In DPPH and ABTS assay, kinnow flavedo showed higher antioxidant capacity that might be due to presence of sufficient TPC and TFC content in kinnow peel while in FRAP assay, orange flavedo showed the highest value. With respect to APCI, kinnow was ranked first followed by orange, malta, hill lemon and mosambi flavedo. Hence, extract from fresh citrus flavedo could be preferred for the utilization in functional food and nutraceuticals as they showed high phytochemical content as well as antioxidant capacity.

REFERENCES

- Abd El-ghfar, MHA., Ibrahim, HM., Hassan, IM., Abdel Fattah, AA. and Mahmoud, MH. (2016). Peels of Lemon and Orange as Value-Added Ingredients: Chemical and Antioxidant Properties. *International Journal of Current Microbiology and Applied Science*, 5(12): 777-794.
- Abu Bakar, M. F., Mohamed, M., Rahmat, A., and Fry, J. (2009). Phytochemicals and antioxidant activity of different parts of bambangan (*Mangifera pajang*) and tarap (*Artocarpus odoratissimus*). Food Chemistry, 113(2): 479–483.
- Agrawal, P.K., Agrawal, C. and Blunden, G. (2021).

 Pharmacological Significance of Hesperidin and Hesperetin, Two Citrus Flavonoids, as Promising Antiviral Compounds for Prophylaxis Against and Combating

- COVID-19. *Natural Product Communications*, 16:1-15.
- Alberca, R.W., Teixeira, F. M. E., Beserra, D. R., de Oliveira, E. A., Andrade, M. M. S., Pietrobon, A. J. and Sato, M. N. (2020). Perspective: The Potential Effects of Naringenin in COVID-19. Frontiers in Immunology, 11: 570919.
- Assefa, A.D., Saini, R.K. and Young-Soo (2017). Fatty acids, tocopherols, phenolic and antioxidant properties of six citrus fruit species: a comparative study. *Journal of Food Measurement and Characterization*. doi:10.1007/s11694-017-9546-x
- Bellavite, P. and Donzelli, A. (2020). Hesperidin and SARS-CoV-2: New Light on the Healthy Function of Citrus Fruits. *Antioxidants* (*Basel*), 9:742.
- Belwal, T., Dhyani, P., Bhatt, I. D., Rawal, R. S. & Pande, V. (2016). Optimization extraction conditions for improving phenolic content and antioxidant activity in Berberis asiatica fruits using response surface methodology (RSM). *Food Chemistry*, 207:115-24.
- Borgonovi, T. F., Virgolin, L. B., Janzantti, N. S., Casarotti, S. N. and Penna, A. L. B. (2022). Fruit bioactive compounds: Effect on lactic acid bacteria and on intestinal microbiota. *Food Research International*, 161:111809.
- Chen, Q., Wang, D., Tan, C., Hu, Y., Sundararajan, B. and Zhou, Z. (2020). Profiling of Flavonoid and Antioxidant Activity of Fruit Tissues from 27 Chinese Local Citrus Cultivars. *Plants*, 9:196.
- Chen, Y., Pan, H., Hao, S., Pan, D., Wang, G. and Yu, W. (2021). Evaluation of phenolic composition and antioxidant properties of different varieties of Chinese citrus. *Food Chemistry*, 364:130413.
- Formagio ASN, Volobuff CRF, Santiago M, Cardoso CAL, Vieira MDC, Valdevina Pereira Z. (2014). Evaluation of Antioxidant Activity, Total Flavonoids, Tannins and Phenolic Compounds in Psychotria Leaf Extracts. *Antioxidants*, 3(4):745-757.
- Gómez-Urios, C., Viñas-Ospino, A., Puchades-

- Colera, P., Blesa, J., López-Malo, D., Frígola, A. and Esteve, M.J. (2023). Choline chloride-based natural deep eutectic solvents for the extraction and stability of phenolic compounds, ascorbic acid, and antioxidant capacity from Citrus sinensis peel. *LWT-Food Science and Technology*, 177:114595.
- Jin, T., YU, M., CAO, M. and Zhu, X. (2022). Optimization of mechanochemical-assisted extraction of hesperidin from Pericarpium Citri Reticulatae. Food Science and Technology. 42:e79821.
- Khan, U. M., Sameen, A., Aadil, R. M., Shahid, M., Sezen, S., Zarrabi, A., Ozdemir, B., Sevindik, M., Kaplan, D. N., Selamoglu, Z., Ydyrys, A., Anitha, T., Kumar, M., Sharifi-Rad, J. and Butnariu, M. (2021). Citrus Genus and Its Waste Utilization: A Review on Health-Promoting Activities and Industrial Application. Evidence-Based Complementary and Alternative Medicine, 2021: 2488804.
- Kim, J.-W., Ko, H. C., Jang, M.-G., Han, S. H., Kim, H. J. and Kim, S.-J. (2023). Phytochemical content and antioxidant activity in eight citrus cultivars grown in Jeju Island according to harvest time. *International Journal of Food Properties*, 26:14-23.
- Li, H.B., Wong, C.C., Cheng, K.W. and Chen, F. (2008). Antioxidant properties in vitro and total phenolic contents in methanol extracts from medicinal plants. *LWT-Food Science and Technology*, 41:385-390.
- Mahato, N., Sinha, M., Sharma, K., Koteswararao, R. and Cho, M. H. (2019). Modern Extraction and Purification Techniques for Obtaining High Purity Food-Grade Bioactive Compounds and Value-Added Co-Products from Citrus Wastes. *Foods*, 8:523.
- Norma Francenia, S.-S., Raúl, S.-C., Claudia, V.-C. and Beatriz, H.-C. (2019). Antioxidant Compounds and Their Antioxidant Mechanism. In: Antioxidants (edited by EMAD, S.). Pp. Ch. 2. Rijeka: IntechOpen.
- Özcan, MM., Ghafoor, K., Al Juhaimi, F., Uslu, N., Babiker, EE., Mohamed Ahmed, IA. and

- Almusallam, IA. (2021). Influence of drying techniques on bioactive properties, phenolic compounds and fatty acid compositions of dried lemon and orange peel powders. *Journal of Food Science and Technology*, 58(1):147-158.
- Saleem. S., Arjumand, I.D., Asnuzilawati, A., Mahmood, A., Muhammad, A., Numan, Y., and Muhammad, M (2023). Investigation of antioxidant and antibacterial effects of citrus fruits peel extracts using different extracting agents: Phytochemical analysis with in silico studies. *Heliyon*, 9(4): e15433.
- Singleton, VL., Orthofer, R. and Lamuela-Raventos, RM. (1999) Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Method Enzymol.*, 299:152–178.
- Rafiq, S., Kaul, R., Sofi, S. A., Bashir, N., Nazir, F. and Ahmad Nayik, G. (2018). Citrus peel as a source of functional ingredient: A review. *Journal of the Saudi Society of Agricultural Sciences*, 17:351-358.
- Rafiq, S., Singh, B. and Gat, Y. (2019). Effect of different drying techniques on chemical composition, color and antioxidant properties of kinnow (*Citrus reticulata*) peel. *Journal of Food Science and Technology*, 56:2458-2466.
- Sharma, K., Mahato, N., Cho, M. H. and Lee, Y. R. (2017). Converting citrus wastes into value-added products: Economic and environmently friendly approaches. *Nutrition*, 34:29-46.
- Siddeeg, A., AlKehayez, N. M., Abu-Hiamed, H. A., Al-Sanea, E. A. and Al-Farga, A. M. (2021). Mode of action and determination of antioxidant activity in the dietary sources: An overview. *Saudi Journal of Biological Sciences*, 28:1633-1644.
- Singanusong, R., Nipornram, S., Tochampa, W. and Rattanatraiwong, P. (2014). Low Power Ultrasound-Assisted Extraction of Phenolic Compounds from Mandarin (Citrus reticulata Blanco cv. Sainampueng) and Lime (Citrus aurantifolia) Peels and the Antioxidant. Food Analytical

Methods, 8:1112–1123.

Tungmunnithum, D., Thongboonyou, A., Pholboon, A. and Yangsabai, A. (2018). Flavonoids and Other Phenolic Compounds from Medicinal

Plants for Pharmaceutical and Medical Aspects: An Overview. *Medicines (Basel)*, 5:93.

Received: March 01, 2024 Accepted: April 02, 2024